PRONTO® ML4 kit
(Mucolipidosis IV)

For the detection of the del(EX1-EX7) and IVS3-2A>G mutations in the MCOLN1 gene

REF **9960**

Instructions for Use

48 **Tests**

PRONTO DIAGNOSTICS

+2°C +8°C IVD CE

INTENDED USE

The PRONTO® ML4 kit is a Single Nucleotide Primer Extension ELISA Assay, intended for the qualitative *in vitro* detection of the del(EX1-EX7) and IVS3-2A>G mutations in the MCOLN1 gene.

For *in vitro* diagnostic use.

BACKGROUND

Similar to some ethnic populations, the Ashkenazi Jewish population has a higher prevalence of certain genetic disorders. These diseases are inherited in an autosomal recessive pattern. Affected individuals have inherited two copies of the mutated gene, one from each parent.
The following table shows the carrier frequency in the Ashkenazi Jewish population and the detection rate of the mutations tested by the PRONTO® ML4 kit.

Disease	Mutation	Carrier Frequency	Disease Frequency	Detection Rate
Mucolipidosis Type IV	IVS3-2A>G del(EX1-EX7)	1/100	1/40,000	95%

REFERENCES

1. Bach G., Mucolipidosis Type IV. *Mol. Genet. Metab.,* 73(3): 197-203. Jul. (2001)
2. Edelman L. et al., Carrier - Screening for Mucolipidosis Type IV in the American Ashkenazi Jewish Population. *Am. J. Hum. Genet.,* 70:1023-1027 (2002).

WARNINGS AND PRECAUTIONS

- Reagents supplied within this kit contain up to 0.1% sodium azide that is very toxic if swallowed. Sodium azide has been reported to form explosive lead or copper azides in laboratory plumbing. To prevent the accumulation of these compounds, flush sink and plumbing with large quantities of water.

- TMB Substrate solution is an irritant material to skin and mucous membranes. Avoid direct contact.
- The Stop Solution contains dilute sulfuric acid (1M), which is an irritant of the eyes and the skin. In case of contact with eyes, immediately flush them with water. Do not add water into this product. In case of an accident or discomfort consult a physician (if possible, present the label).
- In addition to reagents in this kit, the user may come in contact with other harmful chemicals that are not provided, such as ethidium bromide and EDTA. The appropriate manufacturers Material Safety Data Sheets (MSDS) should be consulted prior to the use of these compounds

ASSAY OVERVIEW

The PRONTO® procedure detects pre-defined polymorphisms in DNA sequences, using a single nucleotide primer-extension assay.

Two steps are carried out prior to the use of this PRONTO® kit:

1. **TARGET DNA AMPLIFICATION:** The DNA fragments that encompass the tested mutations are amplified. This amplified DNA is the substrate for the primer extension reaction.
2. **POST-AMPLIFICATION TREATMENT:** The amplified DNA is treated to inactivate free unincorporated nucleotides, so that they will not interfere with the primer extension reaction.
3. **PRIMER EXTENSION REACTION:** carried out in a 96-well thermoplate. Each well contains a 5'-labeled primer that hybridizes to the tested DNA next to the suspected mutation site, and a single biotinylated nucleotide species (corresponding to mutant or wild type), which complements the nucleotide base at the tested site. Each post-amplification treated sample is tested in two wells per mutation: the first well of each pair tests for the presence of the mutant allele (*mut*), while the second well tests for the presence of the normal allele (*wt*). The biotinylated nucleotide will be incorporated in the primer in the course of the reaction or not added, depending on the tested individual's genotype.
4. **DETECTION BY ELISA**: Detection of reaction products is carried out by an ELISA procedure: The biotin-labeled primers bind to a streptavidin-coated ELISA plate and are detected by a peroxidase-labeled antibody conjugate (HRP) directed to the 5' antigenic moiety of the primer. A peroxidase reaction occurs in the presence of the TMB-substrate.
5. **INTERPRETATION OF THE RESULTS**: The results are determined either visually (substrate remains clear or turns blue) or colorimetrically (substrate color remains clear or turns yellow) following the addition of the stop solution.

DISCLAIMER

- Results obtained using this kit should be confirmed by an alternative method.
- Confirmed results should be used and interpreted only in the context of the overall clinical picture. The manufacturer is not responsible for any clinical decisions that are taken.

The user of this kit should emphasize these points when reporting results to the diagnosing clinician or the genetic counselor.

CONTENTS OF THE KIT

2 GeneScreen™ Amplification Mix	1 vial (clear cap)	(13 µL)
PRONTO® Buffer 2	1 bottle	(3mL)
Solution C	1 vial (yellow cap)	(130 µL)
Solution D	1 vial (red cap)	(100 µL)
ColoRed™-Oil	1 dropper bottle	(13 mL)
Assay Solution	1 bottle (green solution)	(100 mL)
Wash Solution (conc. 20x)	1 bottle	(100 mL)
Conjugated HRP	1 vial	(450 µL)
TMB - Substrate	1 bottle	(40mL)
Stop Solution (1M H_2SO_4)	1 bottle	(30 mL)
Detection Plates	2 Streptavidin-coated plates	
PRONTO® ML4 Plates	2 individually pouched plates	

STORAGE AND STABILITY

- Store at 2-8ºC. **Do not freeze.**
- Do not use the kit beyond its expiration date (marked on box label). Stability is maintained even when components are re-opened several times.
- Minimize the time reagents spend at room temperature.
- This kit has been calibrated and tested as a unit; do not mix reagents from kits with different lot numbers.

ADDITIONAL MATERIALS REQUIRED

- Taq DNA polymerase
- Deionized water (about two liters)
- Thermowell plate or tubes (thin walled) for the post-amplification treatment
- Sterile pipette tips
- Troughs/reagent reservoirs - for use with the detection reagents
- Thermocycler for a 96-well microplate
- Multichannel pipettes (5-50 µL and 50-200 µL)
- Positive displacement pipettes (1-5µL, 5-50 µL, 50-200 µL, 200-1,000 µL)
- Filtered tips
- ELISA reader with a 450 nm filter (optional 620 nm filter)
- Polaroid camera and color film to record results (optional)
- Automated microtiter plate washer or a wash bottle
- Vortex mixer
- Timer

ASSAY PROCEDURE

1 DNA AMPLIFICATION

1. **Dispense** 2 µL template DNA (from an initial concentration of about 150 ng/µL) to a thermoplate well or tube.

2. **Prepare** a Master Mix in a sterile vial, according to the volumes indicated in the table below, plus one spare reaction volume. Add the Taq DNA polymerase to the Master Mix shortly before dispensing the Mix. Gently mix by pipetting in and out several times.

PCR Master mix

Solution	Volume for one sample
Amplification Mix 2 GeneScreen™	13.0 µL
Taq DNA polymerase (5 u/µL)	0.5 µL

The following Taq DNA polymerases (lacking 3' → 5' exonuclease activity), were validated for use with this procedure:

- BioLine Cat. # M95801B
- Perkin Elmer Cat. # M801-0060
- Pharmacia Cat. # 27-0799
- ProMega Cat. # M-1661
- Roche Cat. # 1-146-165
- Sigma Cat. # D-1806

3. **Dispense** 13.5 µL Master Mix to each well or tube.

4. **Add** one drop of ColoRed™ oil to each well. Do not touch the wells with the tip of the oil bottle. Even when using a thermocycler with a hot lid, it is recommended to use oil.
5. **Place** the thermoplate well or tube in a thermocycler previously programmed with the following protocol:

Cycling protocol

1.	94ºC	5 minutes	
2.	94ºC	30 seconds	10 cycles
3.	60ºC	60 seconds	
4.	72ºC	60 seconds	
5.	94ºC	30 seconds	25 cycles
6.	55ºC	30 seconds	
7.	72ºC	45 seconds	
8.	72ºC	2 minutes	

6. To verify amplification, **subject** 5 µL of the amplified product to electrophoresis in a 2% agarose gel.

Sizes of amplified fragments:

Gene	Mutations	Fragment size
MCOLN1	Del(ex 1-7)	250 bp
	IVS3-2A→G	386 bp
SMPD1	L302P	515 bp
	fsP330	
	R496L	601 bp
	R608del	

2 POST- AMPLIFICATION TREATMENT

! Only 5 µL of every amplified sample will be used to carry out this assay.

1 **Prepare** a post-amplification treatment mix shortly before use. Combine in a single test tube the volumes appearing in the following table, multiplied by the number of tested samples, plus one spare volume.

Post-Amplification Mix

Solution	Volume for one sample
Amplified DNA	5.0 µL
PRONTO® Buffer 2	45.0 µL
Solution C	2.0 µL
Solution D	1.5 µL

3 **Mix** gently by pipetting this solution in and out five times. Do not vortex.

4 **Add** 48 µL of the post-amplification mix into each well or tube containing **5** µL of each amplified DNA sample.
Ensure that the solution you add becomes well mixed with the DNA sample by inserting the tip under the oil, down to the bottom of the tube and mixing the two solutions by pipetting.

5 **Add** one drop of ColoRed™ oil to each tube.
Do not touch the tube with the tip of the oil bottle.
Even when using a thermocycler with a hot lid, it is essential to use oil.

6 **Incubate** for 30 minutes at 37ºC, then for 10 minutes at 95ºC in a thermocycler.
If not used immediately, the treated sample can be kept at 2-8ºC for a maximum of four hours.

3 PRIMER EXTENSION REACTION

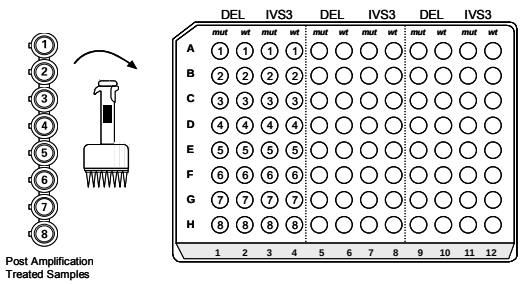
1 **Program** the thermocycler as follows:

Cycle		Temperature	Time
Start:		94ºC	15 sec.
20 cycles:	{	94ºC	30 sec.
	{	63ºC	20 sec.
End:		18-25º C - Cool down to room temperature	

2 **Take** a PRONTO® Plate out of its pouch. Notice the color at the bottom of the wells. For each mutation tested, use a pink well (*mut*) and a blue well (*wt*). Mark the plate with the ID numbers of your test.
If you intend to use less than a full plate, you can cut the plate and return the unused portion to the pouch. If you do this, seal the pouch immediately with its desiccant card inside.

3 **Dispense** 8 µL of post-amplification treated DNA into the first **four** wells in row A (see Fig. 1). Continue with the remaining samples. It is possible to transfer up to eight samples simultaneously using a multichannel pipette.
Ensure that the solution is at the bottom of each well by inspecting the plate from below. Be sure that the well does not contain air bubbles.

Figure 1: Scheme for Dispensing Post-Amplification-Treated DNA Samples into the PRONTO® ML4 Plate.



Recommendation:
Use a new set of tips for each column. Alternatively use the same set of tips, but do not touch the bottom of the wells.

4 **Tilt** the plate and add one drop of ColoRed™ Oil to each well. Do not touch the well with the tip of the oil bottle. Even when using a thermocycler with a hot lid, it is essential to use oil.

5 **Turn on** the thermocycler and start the cycling protocol. Insert the plate when the temperature has reached 90ºC.

6 When the thermal cycling is complete, you can proceed to the ELISA assay, or store the reaction products in the refrigerator and carry out the visualization steps within 24 hours.

4 ELISA ASSAY - COLOR DEVELOPMENT

The ELISA assay consists of the following steps:

- **Binding** the biotinylated primer to the Streptavidin-coated plate.
- **Washing** away the unbound primer.
- **Incubating** with the HRP conjugate.
- **Washing** away the unbound conjugate.
- **Incubating** with the TMB Substrate (color development).

The results of this assay can be determined in one of two ways:

a **Visually**: by monitoring the development of the blue color.

or

b **Colorimetrically:** by adding Stop Solution and measuring the absorbance using an ELISA reader at a wavelength of 450 nm (yellow color)

> **! Before proceeding with the ELISA assay make your choice of visual or colorimetric determination of results.**

PREPARATIONS

- All components used in the detection step should reach room temperature before starting the assay.
- **Dilute** the 20x Wash Solution to 1x with deionized water.
Dilute solution may be kept at 18-25º C for up to one month.

- **Peel** off the plastic cover of the Detection Plate. Mark the side of the plate with the kit name and test number.
- **Place** the PRONTO® plate and the Detection plate side by side, oriented in the same direction (see Fig. 2).

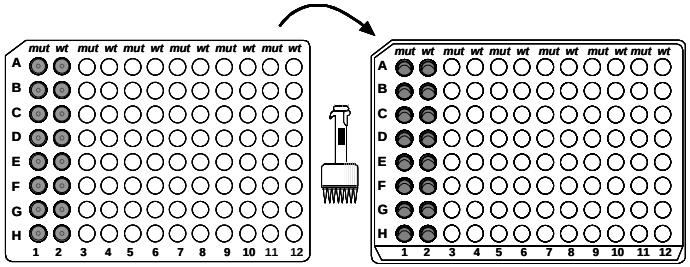
TRANSFER TO THE DETECTION PLATE

1 **Fill** a reagent reservoir /trough with the green colored Assay Solution. About 11 mL will be required for a 96-well plate.

2 **Add** 100 µL of Assay Solution to the bottom of each well in column 1 of the PRONTO® Plate with a multichannel pipette. Mix the Assay Solution with the solution in the wells.

3 **Without changing tips, transfer** 100 µL from each well in this column to the first column in the Detection Plate (see Fig. 2).
Ensure that the solution at the bottom of all wells of the PRONTO® *plate has turned green by inspecting them from below.*

Figure 2: Transferring the Primer Extension Products from the PRONTO® Plate to Detection Plate.



4 **Repeat** this procedure, using a new set of tips for each column. It is essential to maintain the order of the samples.
10 µL of oil carried over or 10 µL of the sample left behind will not affect the detection process significantly.

5 **Incubate** for 10 minutes at room temperature (18-25ºC).

DETECTION BY ELISA

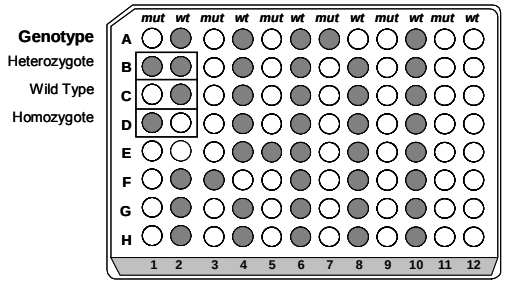
Procedure (continued)		Visual Detection (Blue color)	Colorimetric Detection (Yellow color)
6	While the incubation of Assay Solution is taking place, dilute the Conjugated HRP in Assay Solution For every detection plate (96 wells), use about 11 mL of diluted conjugate **This solution should be freshly prepared each time the test is run.**	Dilution: 1:100 (110 µL of conjugated HRP into 11 mL Assay Solution per plate)	Dilution: 1:250 (44 µL of conjugated HRP into 11 mL Assay Solution per plate)
7	**Empty** the plate and wash four times with 350 µL 1x Wash Solution. Ensure that the plate is dry after the last wash step.	√	√
8	**Add** 100 µL of freshly diluted conjugated HRP to all the wells, with a multichannel pipette.	√	√
9	**Incubate** at room temperature.	10 minutes	10 minutes
10	**Wash** the plate as in step 7.	√	√
11	**Add** 100 µL TMB-Substrate to each well with a multichannel pipette and incubate at room temperature (18-25ºC) until blue color appears	15 minutes	15 minutes
12	**Add** 100 µL of Stop Solution to each well with a multichannel pipette. The solution will turn yellow immediately.	—	100 µL
13	The results can be documented using a Polaroid camera with color film (for example - Fuji FP-100C), or by reading the absorbance using an ELISA reader (signal wavelength setting).	Agitate the plate gently and read results at O.D. 620 nm	—
14	Within two hours **read** the absorbance using an ELISA reader (single wavelength setting).	—	O.D. 450 nm

VALIDATION OF THE RESULTS

For Visual Detection:
For every mutation site tested, at least one of the wells should develop a deep blue color. Otherwise, the results are invalid for the relevant mutation (see Fig. 3).

For Colorimetric Detection:
For every mutation site tested, at least one of the two wells should yield an O.D. ≥ 0.50 reading.

Figure 3: **Visual Interpretation of Genotypes**



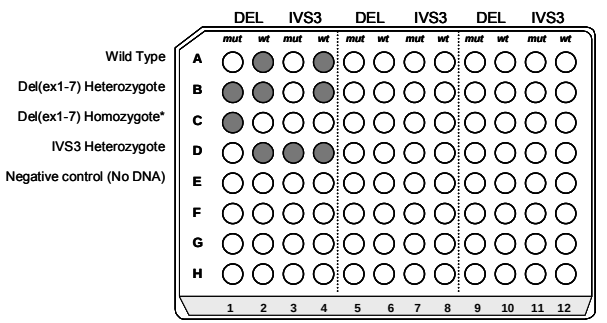
INTERPRETATION OF RESULTS

Important: Heterozygous or homozygous mutant results should be confirmed by retesting. It is recommended to repeat the test with newly extracted DNA.

Criteria for Visual Interpretation

A deep blue color indicates a positive signal, while negative signals appear as a clear or pale blue-colored well (see Fig. 4).

Figure 4: **Examples of genotype assignment according to visual inspection of test results**



* In a del(ex1-7) homozygote sample both wells of the IVS3-2A→G (*mut* and *wt*) should remain clear.

Criteria for Colorimetric Interpretation (O.D. 450)

The genotype of each sample is determined according to two criteria:
1. The O.D. values of the *mut* and *wt* wells.
2. The ratio of *mut* / *wt* O.D. values.

Calculate the *mut* / *wt* ratios by dividing the signal of the *mut* well by the signal of the *wt* well.
Identify the correct genotype using the table below:

Genotype	*mut* well	*wt* well	*mut/wt* ratio
Normal	O.D. ≤ 0.35	O.D. ≥ 0.5	ratio ≤ 0.5
Heterozygote	O.D. ≥ 0.5	O.D. ≥ 0.5	0.5 ≤ ratio ≤ 2.0
Homozygote	O.D. ≥ 0.5	O.D.≤ 0.35	ratio ≥ 2.0

! Samples with values not included in the above table are considered indeterminate and should be retested.

PRONTO® ML4 - PROCEDURE SUMMARY

DNA EXTRACTION from human whole blood using a validated method.

DNA AMPLIFICATION
Volumes per reaction: 2 µL Template DNA + 13.0 µL Amplification Mix + 0.5 µL Taq Polymerase
Cycling protocol: 94°C 5 min→10 cycles of {94°C 30 sec. / 60°C 60 sec./ 72°C 60 sec.} →25 cycles of {94°C 30 sec. / 55°C 30 sec./ 72°C 45 sec.} →72°C 2 min.

POST-AMPLIFICATION TREATMENT:
- Volumes for one reaction

PRONTO® Buffer 2	45.0 µL
Solution C	2.0 µL
Solution D	1.5 µL

- **Pipette** in and out to mix
- **Add** 48 µL into each well containing 5 µL amplified DNA sample and mix well
- **Top** with ColoRed™ Oil.

Incubate 30 minutes at 37ºC, then 10 minutes at 95ºC

PRIMER EXTENSION REACTION:
- **Dispense** 8 µL of each Post-Amplification treated DNA into four wells of the PRONTO® Plate.
- **Top** off with ColoRed™ -Oil.
- **Start** the cycling protocol:
  94ºC 15 sec → 20 cycles of {94ºC 30 sec. / 63ºC 20 sec.} → Cool

Insert the PRONTO® Plate in the thermocycler when the temperature has reached 90ºC

DETECTION:
- **Add** 100 µL Assay Solution to each well in the PRONTO® Plate and mix.
- **Transfer** 100 µL from each well of the PRONTO® Plate to the respective position in the detection plate. Incubate 10 minutes at RT.
- **Empty** the wells and wash four times with 350 µL of 1x Wash Solution.

	Visual Detection	Colorimetric Detection
▪ Add 100 µL of Conjugated HRP to every well and incubate for 10 minutes at room temperature.	Dilution 1:100	Dilution 1:250
▪ Empty the wells and wash four times with 350 µL of 1x Wash Solution.	√	√
▪ Add 100 µL of TMB - Substrate to each well and Incubate at RT for:	15 minutes	15 minutes
▪ Add Stop solution	–	100 µL per well
▪ Results documentation	Photo or O.D. 620 nm	O.D. 450 nm

For troubleshooting guide, please refer to our website:
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